

Effects of chlorella extracellular polymeric substances on the aggregation and stability of TiO, nanoparticles as electrolytes

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ABSTRACT

The enormous application of commercial TiO₂ nanoparticles (TiO₂ NPs) in industrial products will potentially lead to release of TiO₂ NPs in the environment. The aggregation and stability of TiO₂ NPs in natural aquatic environments are essential in assessing their environmental risks. This paper explores the effects of *Chlorella* extracellular polymeric substances (EPS) on the aggregation and stability of TiO₂ NPs as electrolytes. It was found that an increase in NaCl concentration from 0 to 70 mM, the introduction of 5 mg/L EPS better retarded the decline of the absolute zeta potential of TiO₂ NPs also increased with the concentration of EPS. The synergistic effects of steric hindrance and electrostatic repulsion exerted by EPS is considered to stabilize the TiO₂ NPs in NaCl electrolyte medium. However, when using CaCl₂ as the background electrolyte, the polysaccharide groups of EPS can be bridged by Ca²⁺, promoting the aggregation and stability of TiO₂ NPs, which can vary significantly with change in valency and concentration of the electrolyte, making it crucial to understanding the fate and accessing the risks of applying TiO₂ NPs in natural water matrices.

Keywords: Chlorella EPS; TiO, nanoparticle; Electrolyte; Aggregation; Stability

1. Introduction

Due to their unique physicochemical properties, engineered nanoparticles (ENPs) are frequently employed in industrial practices and daily life. It was estimated by Nowack and Bucheli [1] that the productivity of ENPs will go up to 58,000 tons by 2020 based on the employment of nanomaterials in previous years. ENPs are likely to enter the environment via industrial and domestic wastewater, sludge of wastewater treatment plant and waste incineration residues [2].

Among all ENPs, titanium dioxide nanoparticle is one of the most widely used nanomaterials. TiO₂ NPs has many uses, including to fabricate photocatalysts, solar cells, paints, personal care products, and so on. Wastewater from domestic sewage treatment plants usually contains nano- TiO_2 . Keller et al. [3] estimated that the nano- TiO_2 released into sewage system can reach 870–1,000 tons a year in the United States. In recent years, more, and more researchers are focusing on the ecotoxicity of ENPs. For example, Jia et al. [4] evaluated the effects of TiO_2 nanoparticles and multi-walled carbon nanotubes on the freshwater diatoms. It was found that different exposure time and concentration of the nanomaterials will lead to different physiological and biochemical effects on freshwater diatoms. Liu et al. [5] studied the effects of crystalline forms of TiO_2 NPs on the physiological parameters of *Daphnia magna*, and found that anatase TiO_2 NPs may cause more severe damages

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to aquatic organisms at the same bioaccumulation level. Hu et al. [6] reported that TiO_2 NPs can act as an effective carrier for lead to increase its uptake, bioavailability, and toxicity in zebrafish embryos.

ENPs possess more reactive sites than other contaminants due to their small particle sizes and large specific surface area [7–9]. Once released into the environment, ENPs will interact with environmental components such as natural organic matter or inorganic ions [10–12]. Many studies have proved that the bioavailability and toxicity of ENPs to the organism are highly correlated with their particle sizes and colloidal stability. The environmental behavior of ENPs such as aggregation and sedimentation could remarkably affect the fate and ecotoxicity [13–15]. Some researchers have clarified that the electrostatic force and steric hindrance between nanoparticles will change in the presence of natural organic matter and electrolytes, significantly affecting colloidal stability of nanoparticles [16–21] as a result.

Extracellular polymeric substances (EPS) are common components of the natural environmental organic matter. EPS are the mixture of biomacromolecules, produced by microbes during growth and metabolism [22]. As the matrix of biofilms [23], EPS mainly consists of polysaccharides, proteins, small amounts of nucleic acids, and esters [24,25]. In natural water, much of the total extracellular organic substances were secreted by algae, a crucial part of natural organic matter that plays an important role in the aquatic environment. Adeleye et al. [26] studied the effects of EPS on long-term colloidal stability and metal leaching of single-walled carbon nanotubes, which have demonstrated the improved stability of CNTs by EPS in comparison to Suwanee River natural organic matter. Xu and Jiang [27] investigated the stability of ZnO nanoparticles after adsorption of cyanobacterial EPS, and found that the turbidity of ZnO nanoparticles solutions increased with the introduction of EPS, which has demonstrated that the colloidal stability of ZnO nanoparticles had been enhanced. Kroll et al. [28] found that CeO₂ NPs were stabilized by EPS, and the process would not be affected by pH or illumination. Although many studies have shown the effects of EPS on the aggregation and stability of ENPs, the real natural water matrices are more complicated, and the co-effects of electrolytes and EPS on the environmental behavior of NPs lack sufficient research.

Degussa P25 nano-TiO, is commercially available and proven to show good photoactivity, so it has been widely used in laboratory and industries [29]. In this study, typical P25 TiO, NPs were selected as the targeted nanoparticle, and the effects of Chlorella EPS as different electrolytes were investigated. Firstly, zeta potential was determined to study the effects of EPS on the surface charge of TiO, NPs. Secondly, the dynamic light scattering (DLS) was applied to study the aggregation kinetics of TiO₂ NPs in the presence and absence of EPS as the valence and concentration of electrolyte. Finally, the effects of EPS on the stability of TiO₂ NPs as electrolytes were studied through monitoring the absorbance of TiO₂ by using an ultraviolet spectrophotometer for 12 h with different intervals. This study provides an important reference for estimating the fate and transport of ENPs in the natural aquatic environment.

2. Materials and methods

2.1. Preparation of TiO, NPs stock solution

Titanium dioxide (TiO_2) NPs were purchased from Sigma-Aldrich (USA) with >99.5% purity, the primary average particle size was 21 nm. To prepare the nanoparticle suspension, 20 mg of the nanoparticle powder was added to 100 mL of ultrapure water and was sonicated for 30 min. The stock solution was stored at 4°C and was sonicated for 30 min before being used.

2.2. Initial characterization of TiO, NPs

Powder X-ray diffraction (XRD) of the TiO_2 NPs was performed on a Bruker D2 advance diffractometer (Bruker AXS Gmbh, Karlsruhe, Germany). The shape and morphology of TiO₂ NPs were observed through a scanning electron microscopy (SEM, Hitachi S4800, Japan) with selected area electron diffraction. The size of TiO₂ NPs was confirmed by transmission electronic microscopy (TEM, JEM-2100, JEOL, Japan) (Fig. 1). Initially, TiO₂ NPs dispersed in ultrapure water were imaged through transmission electron microscopy (TEM) by depositing of 10 μ L drop on formvar/ carbon-coated copper grids and then drying overnight. After that, micrographs were obtained at 200 keV. The hydrodynamic diameter of the TiO₂ nanoparticle was determined by using dynamic light scattering (Zetasizer Nano-S90, Malvern Instruments Ltd., UK) at 25°C.

2.3. EPS extraction

Chlorella vulgaris (FACHB-1068 in this study was obtained from the freshwater algae culture collection at the Institute of Hydrobiology (FACHB, Wuhan, China), and EPS was extracted from Chlorella by centrifugation [27]. In brief, the purchased algae liquid was transferred to a conical flask containing BG-11 medium. The organisms were cultured under cool white LED lights with a photoperiod of 12:12 h light-dark cycle, at 25°C ± 0.5°C until reaching stationary phase. Cell densities were measured using a hemocytometer. Soluble EPS fraction was harvested at the stationary growth phase and separated via centrifugation of culture solution for 30 min at 4,000 g at 4°C. The supernatant was collected and filtered by 0.22 µm needle filters (Membrane Solutions Company, US) under sterile conditions. To remove the residual of electrolyte from the growth medium and the low molecular-weight metabolites, EPS was dialyzed for 96 h at 4°C using regenerated cellulose tubular membrane of 3,500 Da cut-off (Viskase, US), and the cellulose tubular membrane was pretreated with NaHCO3 and Na2EDTA to remove impurity. The ultrapure water for dialysis was renewed every 24 h, and the EPS after dialysis was stored at 4°C until use. The total organic carbon (TOC) concentration of the EPS solution was measured by a TOC analyzer (TOC-VCPN, Shimadzu, Japan) before the experiment.

2.4. Attenuated total reflection-Fourier transform infrared spectroscopy

The dialyzed *Chlorella* EPS solution was freeze-dried to obtain dried EPS powder. The dried EPS powder was



Fig. 1. SEM (a) and TEM (b) images of TiO₂ NPs.

deposited on a KBr crystal in a Fourier transform infrared (FTIR) spectrometer (L160-8000, PerkinElmer, MA). Each spectrum was scanned 200 times from 4,000 to 400 cm⁻¹ at a resolution of 4 cm⁻¹ [30].

2.5. Aggregation experiment

The change of TiO₂ hydrodynamic diameter (D_{μ}) over time (t) was determined by dynamic light scattering (DLS) (Zetasizer Nano-S90, Malvern Instruments Ltd., UK) operating with a He-Ne laser at the wavelength of 633 nm. The TiO₂ NPs stock solution was sonicated for 30 min before the DLS measurement. The test concentrations of NaCl ranging from 1 to 60 mg/L was set at EPS concentrations of 1, 3, and 5 mg/L. The test concentrations of CaCl, ranging from 0.1 to 10 mg/L was set at EPS concentrations of 1, 3, and 5 mg/L. For each experiment, 1.2 mL of the pre-prepared mixture suspension was introduced into a cuvette (Titan, Shanghai). The concentration of TiO₂ NPs in the DLS vial was maintained at 40 mg/L. The cuvette was vortexed for 1 s right before measurement. The D_{μ} measurement was monitored over a time period from 1 min to 60 min. During the DLS measurements, the scattered light intensity was detected by a photodetector at a scattering angle of 173°, with each autocorrelation function being accumulated over a period of 10 s. All DLS measurements were conducted under 25°C [31].

2.6. Determination of aggregation kinetics

In this study, the aggregation kinetics of TiO₂ particles in early stage could be explained as the primary rate of the increase in D_h with time *t* as tested by DLS. The primary aggregation rate constant (*k*) is proportional to the primary rate of increase in D_h and inversely proportional to the primary concentration of TiO₂ NPs in the suspension, N_0 [32,33]:

$$k \propto \frac{1}{N_0} \left(\frac{dD_h(t)}{dt} \right)_{t \to 0} \tag{1}$$

According to the method proposed by Huangfu et al. [34], the primary increase of the TiO₂ NPs D_h along with time $(dD_h(t)/dt)$ was calculated during a period from aggregation initiation (t_0) to the time when D_h exceeded $1.5D_{h0}$. The value of $dD_h(t)/dt$ was obtained through linear least squares regression analysis.



The particle attachment efficiency α , which was used to quantify the aggregation kinetics of TiO₂ NPs, is determined by normalizing *k* gained under different solution conditions by the k_{fast} obtained under favorable (fast) aggregation conditions [33,34]:

$$\alpha = \frac{k}{k_{\text{fast}}} = \frac{\frac{1}{N_0} \left(\frac{dD_h(t)}{dt} \right)_{t \to 0}}{\left(\frac{dD_h(t)}{dt} \right)_{t \to 0,\text{fast}}}$$
(2)

When the primary TiO₂ concentrations N_0 were maintained constant for the measurements at different ionic strengths, Eq. (2) can be simplified by neglecting N_0 and α could be calculated by normalizing the initial slope of the aggregation profile at a given electrolyte concentration by the initial slope under favorable (fast) aggregation conditions. In order to calculate α in the presence of EPS, $(dD_h(t)/dt)_{t\to 0, fast}$ was obtained from the average values of $(dD_h(t)/dt)_{fast}$ in the diffusion-limited regime (ionic strength above the critical coagulation concentration (CCC)) in the presence of NaCl or CaCl₂.

2.7. Sedimentation experiments

The sedimentation of TiO₂ NPs under various electrolytes and different EPS concentration was studied by UV-vis spectroscopy (752N, Xin Mao, Shanghai). The concentration EPS was set as 0, 1, 3, and 5 mg/L in 5 mM NaCl solution and 0.5 mM CaCl₂ solutions, respectively. TiO₂ stock solution was sonicated for 30 min and then added into the above solution to obtain a mixture which contains 40 mg/L TiO₂ NPs. The UV-vis absorption of the suspension was measured at the wavelength of 378 nm at different time intervals within 12 h. The sedimentation of TiO₂ was expressed by A/A_0 . Where A_0 is the initial absorbency and A is the absorbency recorded at different intervals during the 12 h experimental period.

2.8. Zeta potential measurement

The zeta potentials of TiO_2 and *Chlorella* EPS were determined by a Zeta potential analyzer (Zetasizer Nano ZS,

Malvern, UK). The concentrations of the *Chlorella* EPS were 1 and 5 mg/L, respectively. For ascertaining the effects of ionic strength, the concentration of NaCl or CaCl₂ was set in the range of 0–70 mM and pH was adjusted to 7. All the experiments were repeated three times.

3. Results and discussion

3.1. Characterization of TiO,

The morphology of TiO₂ NPs can be observed in Figs. 1a and b. SEM of TiO₂ NPs showed a near spherical shape without obvious clumps. TEM indicated that the average size of TiO₂ NPs is close to 20 nm, which was in accordance with the size of 21 nm provided by the manufacturer. The hydrodynamic particle size was detected to be 233.6 ± 6.5 nm by DLS, and the TiO₂ surface charge was negative (-3.38 ± 0.11 mV) in neutral aqueous solution. The reason why hydrodynamic particle size of TiO₂ was much larger than the counterpart powder size might be due to the formation of TiO₂ aggregate. The zeta potential of TiO₂ was -7.38 ± 0.5 in neutral solution.

The XRD pattern of TiO₂ NPs is shown in Fig. 2a. The determined characteristic 2 θ values and [hkl] planes were 25.26 [101], 37.85 [004], 47.91 [200], 53.84 [105], and 62.36 [204], which reflect the features of anatase TiO₂. The result were consistent with the standard JCPDs file number and previously published work [35,36].

3.2. Characteristics of EPS

Attenuated total reflection-Fourier transform infrared (ATR-FTIR) was applied to analyze the structure of *Chlorella* EPS, and the wave numbers from 4,400 to 400 cm⁻¹ were recorded. As shown in Fig. 3, notably, the bands at 1,651; 1,574; and 1,385 cm⁻¹ were assigned to the C=O of amide I, N–H of Amide II in protein, and C=O symmetric stretching vibration of –COO– groups. The bands at 1,000–1,132 cm⁻¹

Fig. 2. X-ray diffraction pattern of TiO₂ NPs.

were ascribed to the C–O, C–C stretching vibration, C–O–C and C–O–H deformation vibrations of the polysaccharides structures. The band at 3,400 cm⁻¹ was mainly attributed to –OH in carbohydrates and proteins. The FTIR spectra revealed that the *Chlorella* EPS contains a large number of functional groups that could be absorbed into the surface of the nanoparticle, hence affecting the aggregation behavior of nanoparticles in an aqueous environment.

3.3. Effects of EPS on surface charge of TiO₂ NPs as electrolyte

Surface charge is one of the determining factors in the aggregation and sedimentation behaviors of nanoparticles in aqueous solutions. Electrolytes were generally considered to compress the electric double layer of nanoparticle and lead to significant aggregation. In order to better reveal the effects of Chlorella EPS on the surface charge of TiO₂ NPs in a natural water environment, effects of EPS on the surface charge of TiO, NPs as the function of the concentration of electrolyte was studied. As shown in Fig. 4a, when the EPS concentration was 1 mg/L, with the increase of the NaCl concentrations ranging from 0 to 10 mM, the zeta potential of TiO₂ NPs changed from -22.3 to -5.3 mV. As the concentration of NaCl further increased from 10 to 70 mM, no significant variation of zeta potential could be detected. When the concentration of EPS was 5 mg/L, with the increase of the NaCl concentrations from 0 to 60 mM, the zeta potential of TiO₂ NPs shifted from -22.8 to -11.8 mV. Fig. 4b shows the effect of EPS on the zeta potential of TiO₂ NPs as the function of divalent Ca²⁺. In the presence of 1 mg/L EPS, the zeta potential of TiO, NPs rise from -22.3 to +4.1 mV with the increase of CaCl, concentration from 0 to 50 mM, and the zeta potential of TiO₂ NPs in the 5 mg/L concentration of EPS mixture was -22.8 to +0.8 mV. Similarly, the absolute zeta potential of TiO₂ NPs in the 5 mg/L EPS solution with the same electrolyte concentration was significantly higher than that of TiO, NPs in the 1 mg/L EPS solution. When the concentration of CaCl, was



Fig. 3. FTIR spectra of EPS.





Fig. 4. Zeta potential of TiO, NPs in the presence of EPS as the function of (a) NaCl and (b) CaCl, concentration.

20 mM, it was obvious that the absolute zeta potential of TiO_2 NPs in 5 mg/L EPS solution was higher than that in 1 mg/L EPS solution. This phenomenon may be associated with the fact that Ca²⁺ neutralizes the negatively charged group of EPS and the surface charge of TiO₂ NPs. Therefore, the addition of *Chlorella* EPS in CaCl₂ solution promotes more aggregation of TiO₂ NPs than that in NaCl solution.

3.4. Effects of EPS on aggregation of TiO, NPs as electrolyte

The electrolyte concentration at the intersection of the reaction/diffusion-limited regimes are defined as CCC [32]. The stability of TiO, NPs in aquatic systems can be predicted by CCC. To explore the effects of EPS on TiO, NPs aggregation rate, the effects of different concentrations of EPS on the aggregation rate of TiO₂ NPs were investigated. With a background of low electrolyte concentrations, the increase in electrolyte concentration will promote the degree of charge screening and lead to an improvement in aggregation kinetics, as reflected by the rise in attachment efficiency, which was known as the reaction-limited regime ($\alpha < 1$). Under higher electrolyte concentrations, the surface charge of TiO, nanoparticles was almost neutralized and the energy barrier between TiO₂ nanoparticles was cleared. Therefore, the diffusion-limited aggregation $(\alpha = 1)$ of nanoparticles is likely to take place. In this situation, aggregation kinetics will reach the maximum and will not be affected by the electrolyte concentration anymore.

The effects of EPS on the attachment efficiencies (α) of TiO₂ nanoparticles as electrolyte (NaCl and CaCl₂) solutions were shown in Fig. 5. As illustrated in Fig. 5a, in the absence of EPS, the CCC of TiO₂ NPs was 11.07 mM in NaCl solution. After the introduction of EPS with concentrations of 1, 3, and 5 mg/L, the CCC of TiO₂ NPs were determined to be 2.79, 14.99, and 23.16 mM, respectively. One milligram per liter EPS promoted the aggregation of TiO₂ NPs in NaCl solution. The point of zero charge (PZC) of TiO₂ NPs was reported to be approximately 6.2 [37], and the surface charge was 7.38 when such kind of TiO₂ NPs appeared in neutral solution. As demonstrated in Fig. 5a, when the

concentration of EPS was 1 mg/L, the surface charge of TiO_2 NPs rapidly decreased with the increase in the concentration of NaCl, which was even lower than that of TiO_2 NPs in the absence of EPS, diminishing the stability of the system as a result. Despite the steric hindrance effects of EPS, the low concentration of EPS could not compensate for the electric repulsion loss caused by the neutralization of surface charge of TiO_2 NPs. The retarding effects are primarily attributed to the electrostatic repulsion. The EPS molecules absorbed on the surface of TiO_2 NPs would slow down the decline of the surface charge of TiO_2 NPs, and this phenomenon could be explained by the enhancement of the electrostatic repulsion among these NPs. Apart from this, EPS attached to the surface also provided spatial hindrance to alleviate the nanoparticles from aggregation.

As shown in Fig. 5b, the CCC of TiO, NPs in CaCl, solution was 3.07 mM in the absence of EPS. After the introduction of EPS with concentrations of 1, 3, and 5 mg/L, the CCC of TiO₂ NPs changed to 0.66, 1.60, and 1.69 mM, respectively. The above result indicated that the introduction of EPS promoted the aggregation of TiO₂ NPs in CaCl₂ containing matrices. This phenomenon may be attributed to the fact that the polysaccharide components of Chlorella EPS contain carboxylic groups, which could crosslink with Ca2+ through intermolecular reaction. Based on this, it can be inferred that in the presence of Chlorella EPS, the increase of the aggregation rate of TiO₂ NPs was caused by the intermolecular bridging between Ca²⁺ and the COO⁻ groups in the polysaccharide. Ca2+ complexes polysaccharide macromolecules can promote the aggregation of TiO₂ NPs, thereby speed up the growth of total agglomerates. It also has been reported that Ca2+ can cause aggregation of polysaccharides by intermolecular bridging of polysaccharides, which further helps to increase the polymerization of it [32-34,38,39].

3.5. Effect of EPS on stability of TiO₂ NPs as the function of electrolytes

In this section, the effects of EPS on the sedimentation behaviors of TiO₂ NPs in the electrolyte containing



Fig. 5. CCC of TiO₂ colloids in (a) NaCl solutions and (b) CaCl₂ solutions.



Fig. 6. Sedimentation of TiO₂ NPs in (a) 5 mM NaCl and 0.5 mM (b) CaCl₂.

matrices were explored. As shown in Fig. 6a, EPS remarkably retarded the sedimentation of TiO_2 NPs in an aqueous matrix containing 5 mM NaCl, and the increase rate of the retarding effect was highly correlated with the concentration of EPS. In the presence of 5 mg/L EPS, 27.0% of the TiO₂ NPs were found suspended after 12 h, 19.8% higher than the counterpart without the introduction of EPS. In NaCl solution, the EPS adsorbed on the surface of TiO₂ NPs resulted in steric hindrance, and the steric hindrance effect would enhance with the increase of EPS concentration, which impeded the sedimentation of TiO₂ NPs.

Nevertheless, as portrayed in Fig. 6b, the coexistence of EPS and divalent Ca²⁺ promoted the sedimentation of TiO₂ NPs. This result was consistent with the aggregation study, the presence of divalent Ca²⁺ bridged the EPS-covered TiO₂ nanoparticles, thereby increasing the formation of large aggregate and promoted the sedimentation process.

4. Conclusion

The effects of Chlorella on the aggregation behavior of TiO₂ nanoparticles were studied in the presence of different electrolytes. With regard to NaCl, the aggregation and sedimentation of TiO₂ NPs were retarded when Chlorella EPS concentrations were 3 or 5 mg/L. The retarding effect of nanoparticle aggregation can be attributed to the electrostatic repulsion and steric hindrance caused by EPS attached to its' surface. In comparison, when divalent Ca²⁺ was selected as the background electrolyte, the introduction of 1-5 mg/L Chlorella EPS promoted the aggregation and sedimentation behavior of TiO₂ NPs. This is because the EPS attached to the surface of nanoparticles contain rich polysaccharides, which can be crosslinked by Ca²⁺. The results indicated that *chlorella* EPS has a great influence on the aggregation and sedimentation process of nanoparticles as electrolytes. This study provides an

important reference for assessing the fate and transport of ENPs in real natural waters. However, the water chemistry of natural aqueous environment is more complicated. To understand the fate of more nanoparticles in actual aquatic environment, further researches on the behaviors of ENPs in natural waters are still needed.

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